

FIG 1. The balance between nutrition and malnutrition and the factors that preponderate on the outcome of health and disease in this interplay are shown.

Balneoterapie – imunomodulační nástroj



Doplňkové léčebné možnosti (as.MUDr.Š.Čapková)

- **Lázeňská léčba:** klimaticky vhodná oblast, koupele, léčba dermatologická
- **Přímořská léčba:** fototerapie, vliv podnebí, sůl na kůži
- **Klimatická léčba (hory):** nízké množství alergenů, chladnější klima



Balneoterapie – imunomodulační nástroj

- ✓ UV terapie
- ✓ modulace nespecifické imunity
- ✓ změna prostředí
- ✓ stres

Balneotherapie – immunomodulační nástroj

✓ UV terapie

■ **Tabelle 106.1.** Einteilung der optischen Strahlung

| Wellenbereich | Abkürzung | Wellenlänge ^a |
|-------------------------|-----------|--------------------------|
| Kurzwelliges UV | UVC | 200–280 nm |
| Mittelwelliges UV | UVB | 280–320 nm |
| Langwelliges UV | UVA | 320–400 nm |
| | UVA2 | 320–340 nm |
| | UVA1 | 340–400 nm |
| Sichtbares Licht | SL | 400–800 nm |
| Kurzwelliges Infrarot | IR-A | 800–3000 nm |
| Mittelwelliges Infrarot | IR-B | 3–10 µm |
| Langwelliges Infrarot | IR-C | 10 µm–1 mm |

^a International in den Naturwissenschaften gebräuchliche Einteilung. Daneben bestehen in nationalen Richtlinien geringfügig abweichende Definitionen (wie nach DIN: UVB: 290–315 nm, UVA: 315–380 nm)

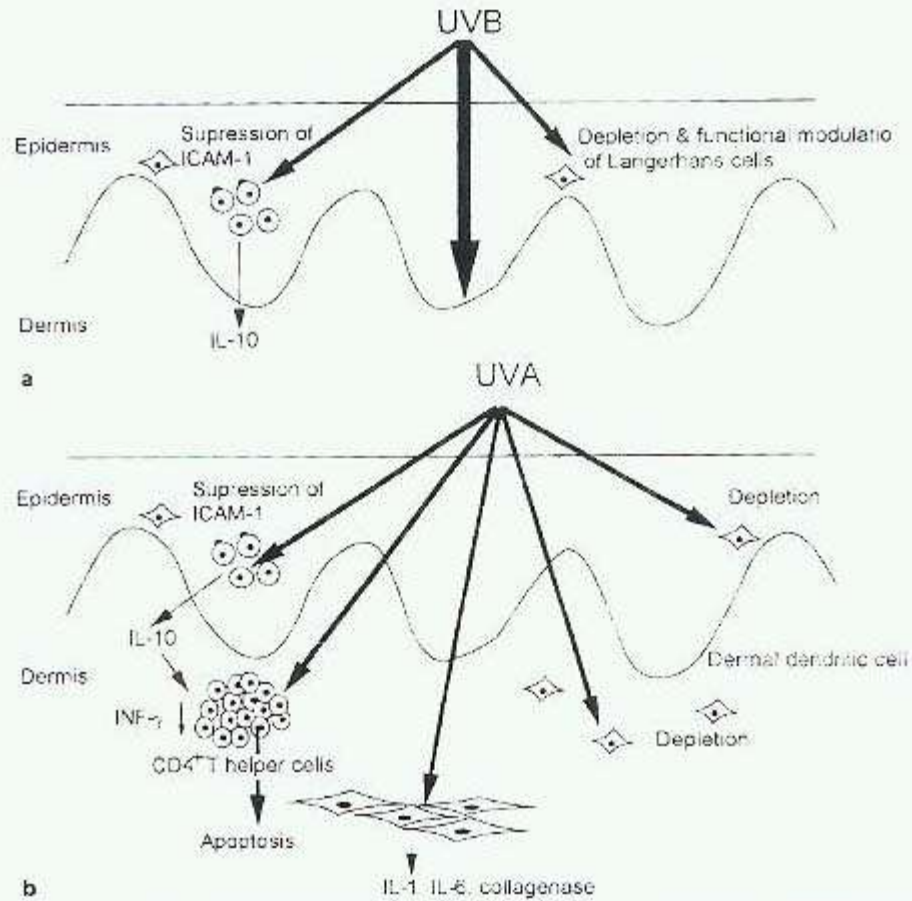


Fig. 1a, b. Scheme of immunomodulatory effects induced by UVB- [A] or UVA(i)-[B] photo(chemo)therapy. *ICAM-1*, intercellular adhesion molecule-1; *INF- γ* , interferon- γ ; *IL-10*, interleukin-10

Table 1. Subsets of natural and induced regulatory T cells¹

| Treg subset | Regulatory mechanisms | Transcription factor expressed | Target cells | Function |
|--|---|--------------------------------|----------------------------|--|
| CD4 ⁺ CD25 ⁺ Tregs | Cell contact-dependent, cytokines (IL-10 [?]) | Foxp3 | T cells, APCs | Suppression of autoimmunity; inhibition of allograft rejection and of immune responses induced by microbial infection; mediation of UV-induced immunosuppression |
| CD4 ⁺ CD25 ⁻ Tregs | Mostly mediated by cytokines | Foxp3 (??) | T/B cells, APCs | Suppression of autoimmunity |
| Tr1 cells | Mediated by IL-10 | Foxp3 (??) | T cells | Suppression of autoimmunity |
| Th3 cells | Mediated by TGF- β | ?? | T cells | Suppression of autoimmunity |
| NKTregs | IL-4, IL-10, TGF- β , cytotoxicity | ?? | T cells, APCs, tumor cells | Elimination of tumors and pathogens; suppression of autoimmunity; mediation of UV-induced suppression of protective tumor immunity |
| CD8 ⁺ Tregs | Cell contact-dependent, cytotoxicity, cytokines (??) | Foxp3 (??) | T cells | Suppression of autoimmunity; regulation of peripheral TCR repertoire |
| CD8 ⁺ CD28 ⁻ Tregs | Induction of ILT3/ILT4 in DCs | Foxp3 (??) | DCs/APCs | Regulation of autoimmunity (?? ^b) |

¹Subsets have been detected in humans and rodents. ²Issue uncertain, not yet clear or not yet investigated. Abbreviations: APC, antigen-presenting cell; DC, dendritic cell; ILT, immunoglobulin transcript; NKTreg, regulatory cell of natural killer T cell phenotype; Th3, T helper type 3; Tr1 cell, type 1 regulatory T cell; Treg, regulatory T cell.

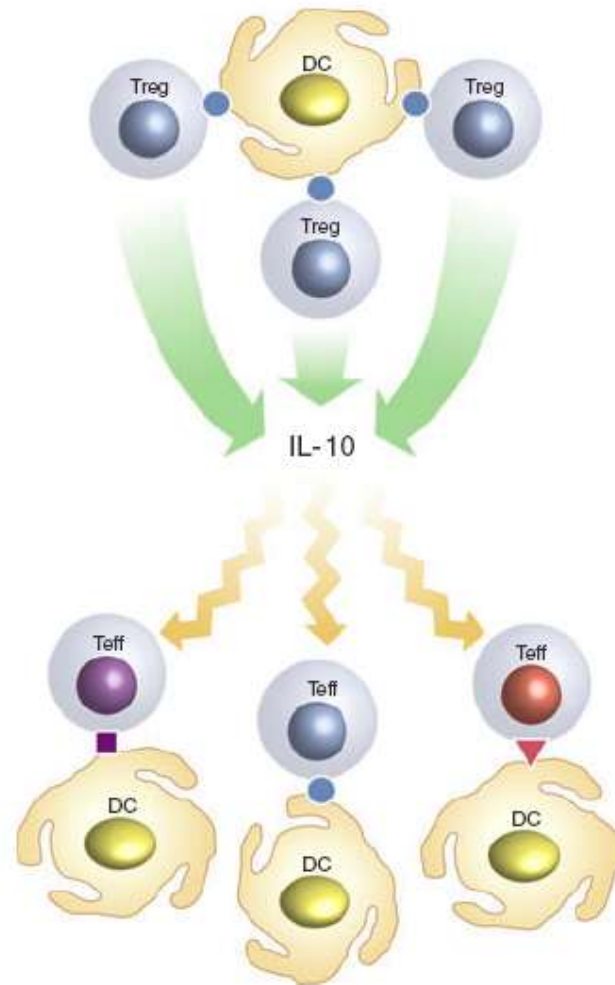
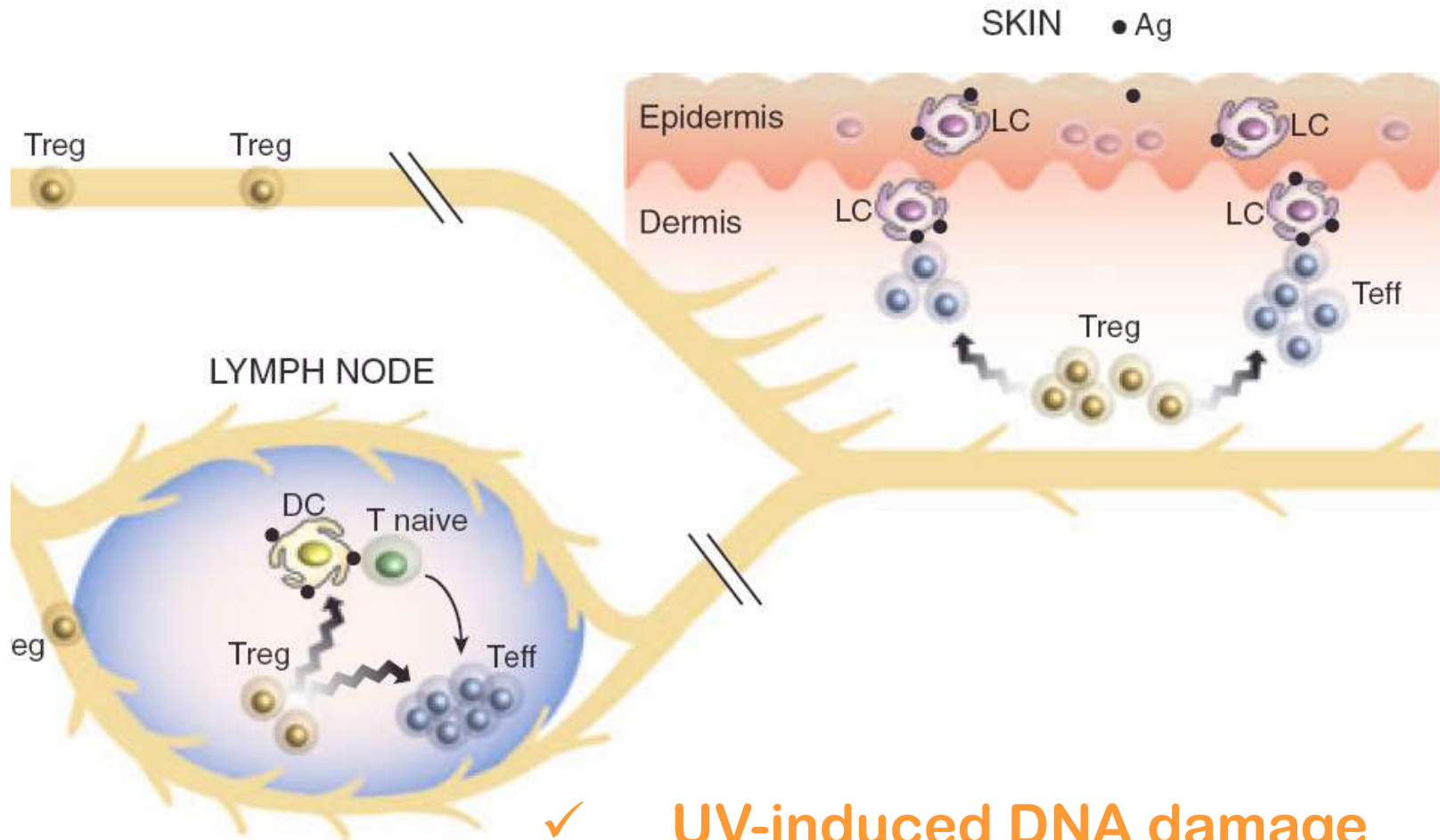


Figure 2. Mechanism of bystander suppression. Tregs that are activated by DCs in an antigen-specific fashion release IL-10. Once released, IL-10 inhibits immune reactions not only against the initial antigen (●) but also against other antigens (▼, ■) in a nonspecific fashion.



- ✓ UV-induced DNA damage
- ✓ IL-10 Treg
- ✓ IL-12 reduces LC-DNA damage

Solar-Simulated Ultraviolet Radiation Induces Abnormal Maturation and Defective Chemotaxis of Dendritic Cells

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Exposure to ultraviolet (UV) light induces immunosuppression. Different evidences indicate that this phenomenon is mainly a consequence of the effect of UV light on skin dendritic cells (DC). To investigate the cellular and molecular basis of this type of immunosuppression, we assessed *in vitro* the effect of solar-simulated UV radiation on the phenotypic and functional characteristics of human monocyte-derived DC and Langerhans-like DC. UV radiation induced a decreased expression of molecules involved in antigen capture as DC-SIGN and the mannose receptor. This effect was accompanied by a diminished endocytic capacity, an enhanced expression of molecules involved in antigen presentation such as major histocompatibility complex-II and CD86, and a significant increase in their capability to stimulate T cells. Furthermore, irradiated DC failed to acquire a full mature phenotype upon treatment with lipopolysaccharide. On the other hand, solar-simulated radiation induced the secretion of tumor necrosis factor- α and interleukin (IL)-10 by DC, but no IL-12. Interestingly, solar-simulated UV radiation also caused an altered migratory phenotype, with an increased expression of CXCR4, and a lack of induction of CCR7, thus correlating with a high chemotactic response to stromal cell-derived factor 1 (SDF-1) (CXCL12), but not to secondary lymphoid tissue chemokine (SLC) (CCL21). These data indicate that solar-simulated UV radiation induces a defective maturation and an anomalous migratory phenotype of DC.

Key words: chemotaxis/costimulation/cytokines/dendritic cells/human
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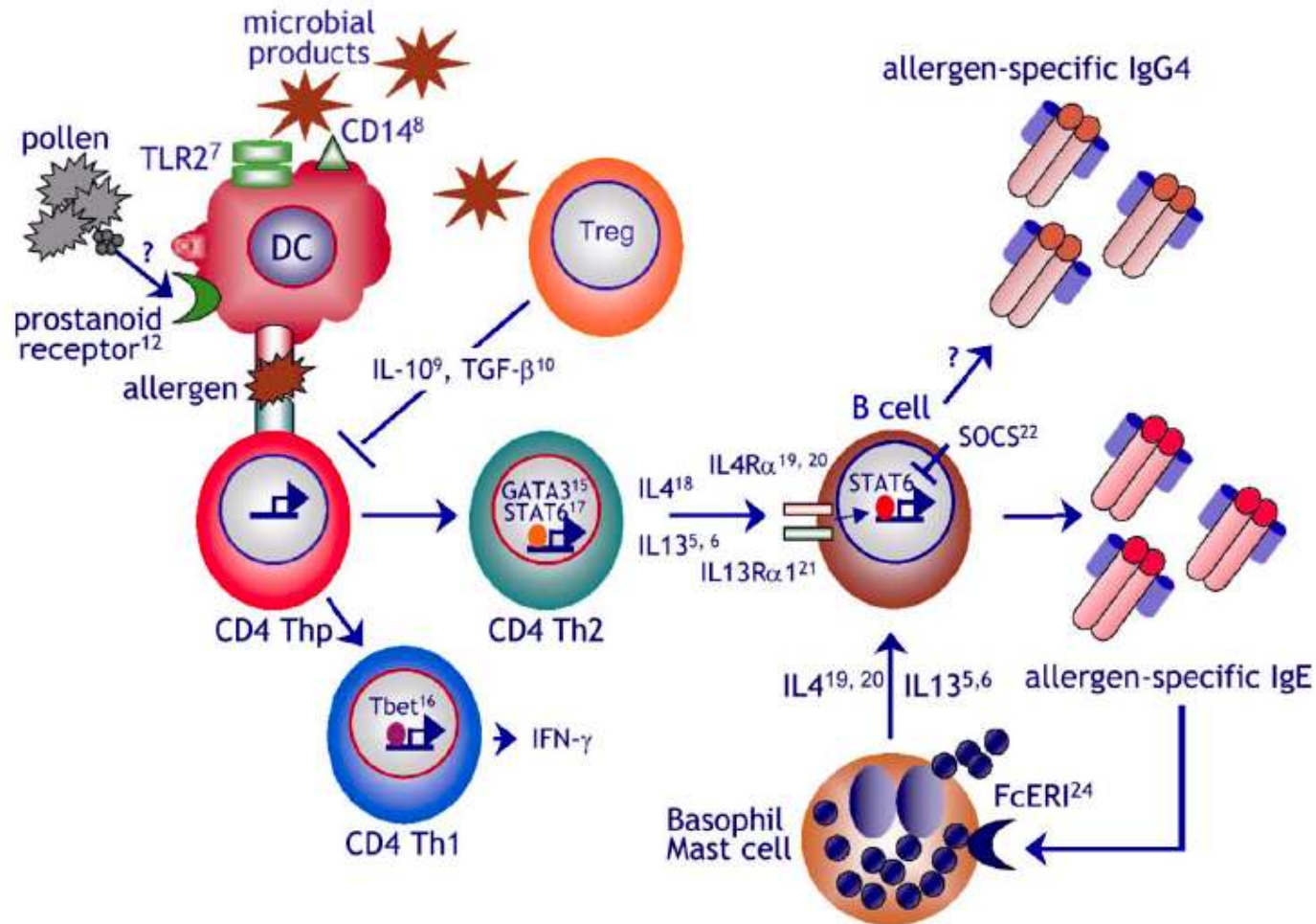


FIG 1. Gene-gene interactions and the pathogenesis of allergic inflammation: a working roadmap. Groups of interactions are color coded: the regulatory and sensing interfaces are in red/orange, T_H differentiation is in blue/green, and the effector phase is in shades of brown. See text for an in-depth discussion of individual pathways and genes. References are numbered as in the text.

■ **Tabelle 106.5.** Indikationen zur UVB-Phototherapie

| Indikation | UVB-Breitband | UVB-311 nm |
|---|---------------|------------|
| Psoriasis | + | ++ |
| Atopisches Ekzem | + | ++ |
| Pruritus, Prurigo | + | + |
| Parapsoriasis en plaques | + | + |
| Mycosis fungoides (Patch-Stadium) | + | + |
| Prophylaxe der polymorphen Lichtdermatose | + | ++ |
| Vitiligo | - | ++ |
| Pityriasis lichenoides | + | 0 |
| Lymphomatoide Papulose | + | 0 |
| Seborrhoisches Ekzem | + | + |
| HIV-assoziierte pruritische Eruptionen | + | 0 |

+ empfehlenswert, ++ Überlegen, - gering wirksam, 0 keine Erfahrungsberichte

■ **Tabelle 106.8.** Indikationen für UVA1-Phototherapie

| Standardtherapie | Wissenschaftliche Erprobung |
|-------------------------------------|---|
| Atopisches Ekzem | Urticaria pigmentosa |
| Dyshydrosiformes Hand- und Fußekzem | Granuloma anulare |
| Morphea | Sarkoidose |
| | Lichen sclerosus et atrophicus |
| | Lichturtikaria |
| | Mycosis fungoides |
| | Psoriasis |
| | Pruritus |
| | Prurigo |
| | Akrale systemische Sklerodermie |
| | Akute und chronische Graft-versus-Host-Erkrankung |

■ Tabelle 106.10. Häufigste Indikationen für Photochemotherapie

| Diagnose | PUVA oral | PUVA-Vollbad | PUVA-Hand- und Fußbad |
|--|-----------|----------------|-----------------------|
| Psoriasis vulgaris | + | + | |
| Palmoplantare Psoriasis | | | + |
| Atopisches Ekzem | + | + | |
| Dyshidrosiformes und hyperkeratotisches Hand- und Fußekzem | | | + |
| Parapsoriasis en plaques | + | + | |
| Mycosis fungoides | + | + ² | |
| Lymphomatoide Papulose | + | + | |
| Morphea | + | + | |
| Akute und chronische (sklerodermiforme) Graft-versus-Host-Erkrankung | + | + | |
| Lichen ruber | + | + | + |
| Photodermatosen | + | | |
| Polymorphe Lichtdermatose | | | |
| Lichturtikaria | | | |
| Chronische aktinische Dermatitis | | | |
| Hydroa vacciniformia | | | |
| Aktinische Prurigo | | | |

² Gesicht muss von der Behandlung ausgespart bleiben, daher kann nicht die bei Mycosis fungoides meist notwendige Ganzkörpertherapie erfolgen

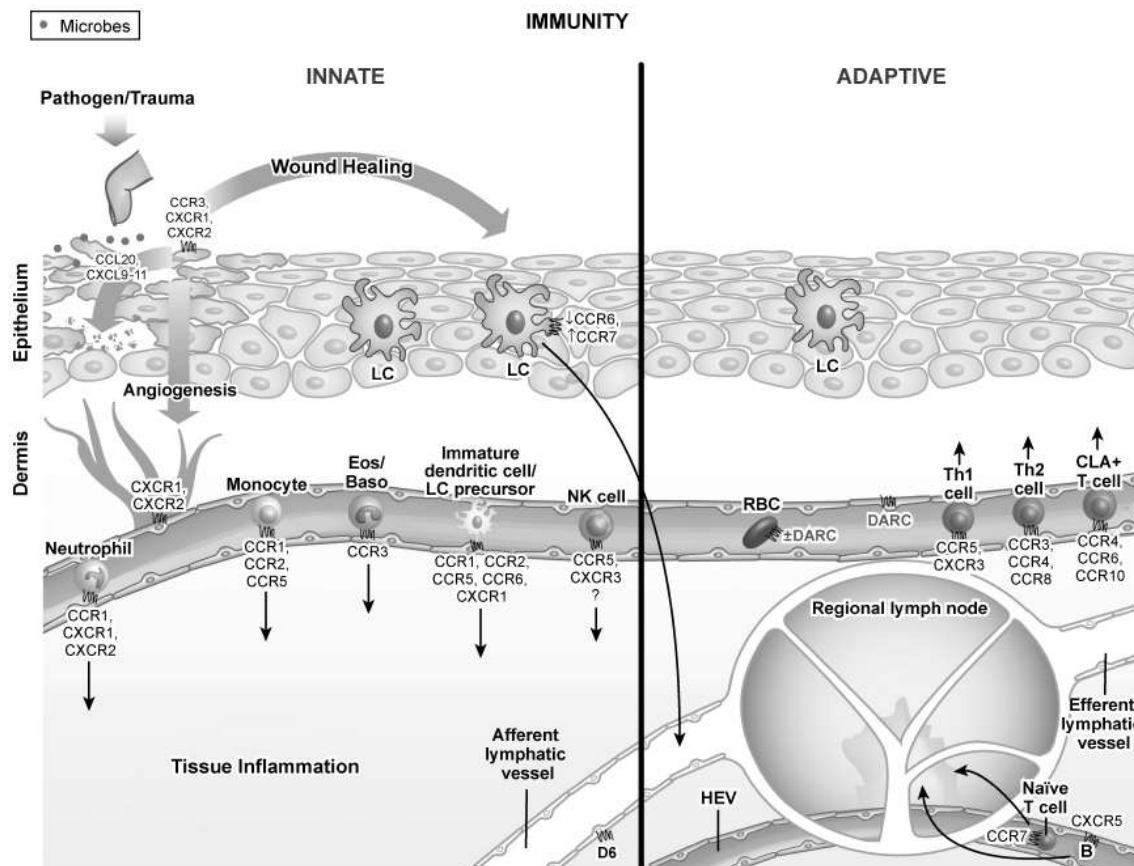
■ **Tabelle 106.14. Risiken und Nebenwirkungen der Photo(chemo)therapie**

| | UVB | UVA/UVA1 <20 J/cm ² | UVA1 >20 J/cm ² | PUVA |
|--|-----|--------------------------------|----------------------------|------|
| Sonnenbrand und phototoxische Reaktion bei Überdosierung | ++ | - | - | ++ |
| Phototoxische Reaktion durch unbeabsichtigte Zufuhr eines Photosensibilisators | ± | + | ++ | ++ |
| Konjunktivitis und Keratitis bei fehlendem Augenschutz | ++ | - | - | ++ |
| Provokation von Photodermatose (polymorphe Lichtdermatose) | + | + | ++ | ± |
| UV-Lentigines | + | ± | + | ++ |
| Lichtalterung der Haut | ++ | ± | ++ | ++ |
| Präkanzerosen und spinozelluläres Karzinom | + | ? | ± | ++ |
| Melanome | ? | ? | ? | ? |

++ hohes Risiko; + mäßiges Risiko; ± geringes Risiko; - aufgrund bekannter Wirkmechanismen nicht wahrscheinlich; ? prinzipiell möglich, aber keine Daten vorhanden

Balneoterapie – imunomodulační nástroj

✓ modulace nespecifické imunity



Humorální faktory v kožním imunitním systému

Antimikrobiální peptidy -defensiny, cathelicidiny, dermcidiny

Lysozym

Složky komplementového systému (C3, fB, fH; CD59 /DAF/, CD46 /MCP/, CD59; CR1, CR2)

Cytokiny (IL-1, TNF- α , ... interferony, chemokiny...)

Imunoglobuliny (IgG, IgA vč. SIgA)

Fibrinolysiny

Produkty koagulační kaskády

Eikosanoidy a prostaglandiny

Neuropeptidy

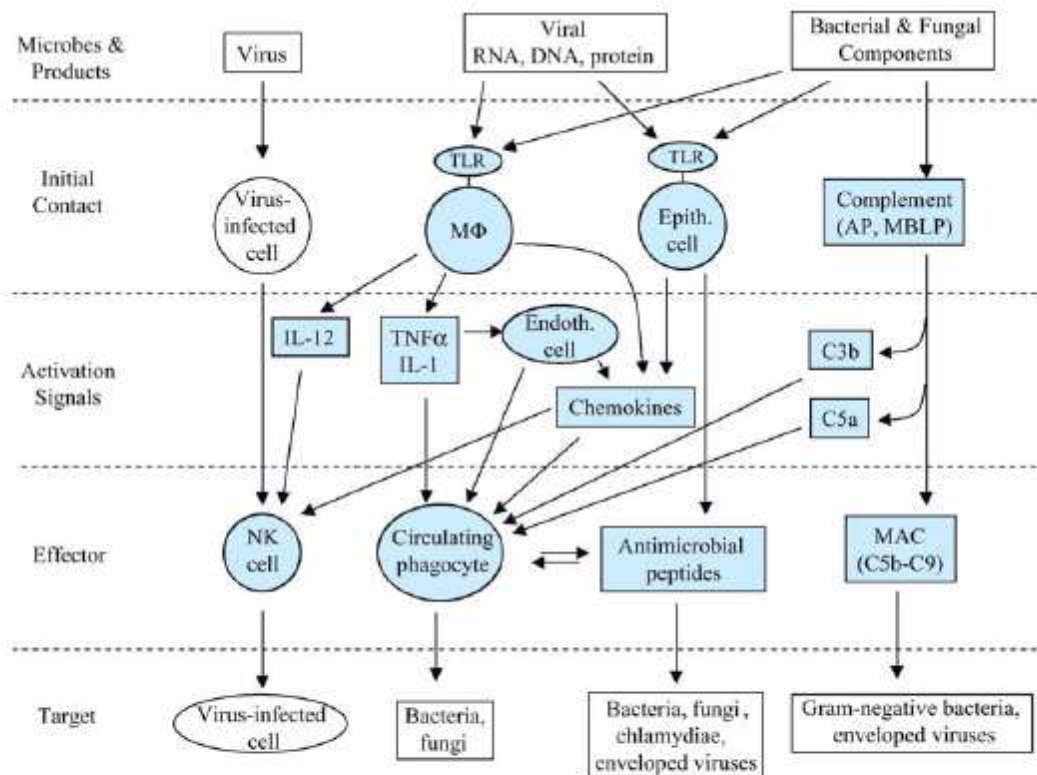


FIG 1. Innate immunity: responses to first contact. Diagrammed are important host responses to infection that are independent of specific cell-mediated immunity or antibodies. Initial contact between the host and microbes or their products results in a range of activating signals that mobilize both cellular and humoral effectors for attack on their respective microbial targets. Components of the host response are highlighted in blue. *MΦ*, Macrophages; *AP*, alternative pathway; *MBLP*, mannose-binding lectin pathway.

Stratum Corneum Defensive Functions: An Integrated View

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Most epidermal functions can be considered as protective, or more specifically, as defensive in nature. Yet, the term “barrier function” is often used synonymously with only one such defensive function, though arguably its most important, i.e., permeability barrier homeostasis. Regardless of their relative importance, these protective cutaneous functions largely reside in the stratum corneum (SC). In this review, I first explore the ways in which the multiple defensive functions of the SC are linked and interrelated, either by their shared localization or by common biochemical processes; how they are co-regulated in response to specific stressors; and how alterations in one defensive function impact other protective functions. Then, the structural and biochemical basis for these defensive functions is reviewed, including metabolic responses and signaling mechanisms of barrier homeostasis. Finally, the clinical consequences and therapeutic implications of this integrated perspective are provided.

Key words: barrier function/corneodesmosomes/cytokines/desquamation/hydration/lamellar bodies/pH/psychological stress/stratum corneum/lamellar body

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Table I. Protective functions of mammalian stratum corneum

| Function | Localization |
|---|---------------------|
| Permeability barrier ^a | Extracellular |
| Cohesion (integrity) → desquamation ^a | Extracellular |
| Antimicrobial barrier (innate immunity) ^a | Extracellular |
| Mechanical (impact and shear resistance) | Corneocyte |
| Toxic chemical/antigen exclusion | Extracellular |
| Selective chemical absorption | Extracellular |
| Hydration | Corneocyte |
| UV barrier | Corneocyte |
| Initiation of inflammation (cytokine activation) ^a | Corneocyte |
| Psychosensory interface | Unknown |
| Thermal barrier | Unknown |

^aRegulated or thought to be activated by (SC) pH.

STRATUM GRANULOSUM (SG)

Lipid Precursors

glucosylceramides,
cholesterol,
glycerophospholipids,
sphingomyelin



Catabolic Enzymes

serine proteases, lipases

Other

corneodesmosin,
beta-defensin 2,
acid phosphatase,
glycosidases,
protease inhibitors

SG-STRATUM CORNEUM INTERFACE

Conversion Into Non-polar Lipid Products

(lipases, glycosidases)

Glucosylceramides → Ceramides 1-7 →

Sphingomyelin → Ceramides 2,5 →

Phospholipids → FFA →

Cholesterol →



1) **Degradation** of corneodesmosomes (serine proteases)

2) **Degradation** of other extracellular species (acid phosphatase, glycosidases)

LOWER STRATUM CORNEUM

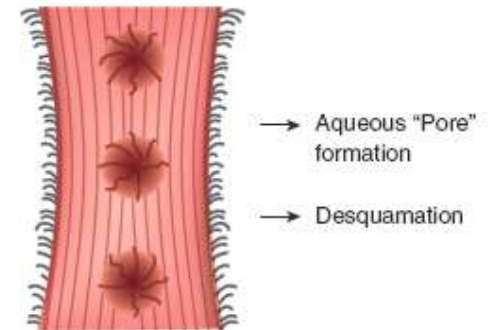
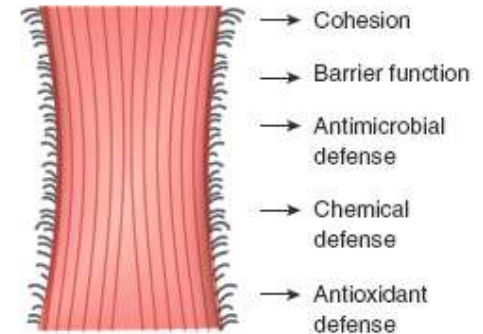


Figure 1

Lamellar body secretion dictates localization of multiple functions to extracellular compartment.

Hydration, ultraviolet (UV) filtration, and immunosuppression are linked through the histidase pathway. In addition to resistance of the skin to mechanical or blunt injury, cornocytes ("bricks") generate filaggrin-derived peptides and their deiminated products, which, along with sebaceous gland-derived glycerol (Fluhr *et al.*, 2003), regulate not only SC hydration but also several other downstream functions (Fig 2). Early in cornification, filaggrin, the predominant, histidine-enriched, basic protein in F-type keratohyalin granules, disperses around keratin filaments within the stratum compactum (Dale *et al.*, 1997). At ambient humidities (<85% RH), i.e., above the stratum compactum, filaggrin is largely hydrolyzed by a still-uncharacterized, cytosolic protease into free amino acids, including histidine, glutamine (glutamic acid), and arginine (Scott and Harding, 1986; Harding *et al.*, 2000). These amino acids, and their distal, deiminated products (urocanic acid, pyrrolidone carboxylic acid, and ornithine/citrulline/aspartic acid, respectively) comprise much of the osmotically active material that regulates SC hydration (Harding *et al.*, 2000). Histidine is deiminated enzymatically to its acidic, polar, plurifunctional metabolite, trans-urocanic acid (tUCA), by the enzyme, histidine ammonia lyase (histidase) (Scott, 1981). As it acts as an *endogenous sunscreen*, tUCA is photoisomerized by UV-B to cUCA (De Simone *et al.*, 2001), a potent *immunosuppressive* molecule, implicated in the pathogenesis of UV-induced skin cancers (Noonan and De

cohesion by a common mechanism, i.e., suppression of epidermal lipid synthesis and LB production (Kao *et al.*, 2003). Accordingly, not only the stress-induced barrier abnormality, but also the abnormality in SC integrity can be reversed (overridden) by a mixture of physiologic lipids, containing all three key SC species (i.e., ceramides, FFA, and cholesterol) (Kao *et al.*, 2003).

Impact of pH on multiple defensive functions A second example of a common stressor that modulates multiple functions is SC pH. In fact, pH orchestrates at least three important SC defensive functions (Table I). Using a flat surface electrode, the pH of mammalian SC typically ranges from 4.5 to 5.0 in the outer SC, approaching neutrality in the lower SC (Ohman and Vahlquist, 1994). Recent studies, utilizing fluorescence-lifetime imaging (FLIM) combined with multiphoton microscopy, show that SC pH is heterogeneously distributed, and that the pH gradient is non-linear (Behne *et al.*, 2002, 2003). Most importantly, FLIM demonstrated that membrane domains even at the level of the stratum granulosum (SG)-SC interface are selectively acidified (Behne *et al.*, 2002).

Although at least two endogenous, biochemical pathways regulate SC pH, the full array of functions that are impacted by each mechanism is still largely unclear. A third mechanism is the histidine-to-UCA pathway, which can account quantitatively for SC acidification (Krien and

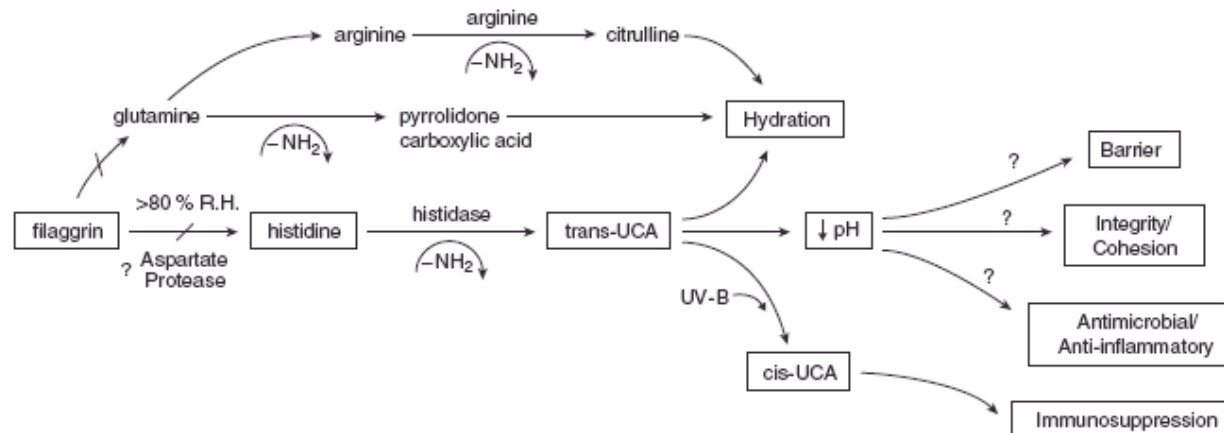


Figure 2
Functions potentially impacted by filaggrin metabolism in the stratum corneum.

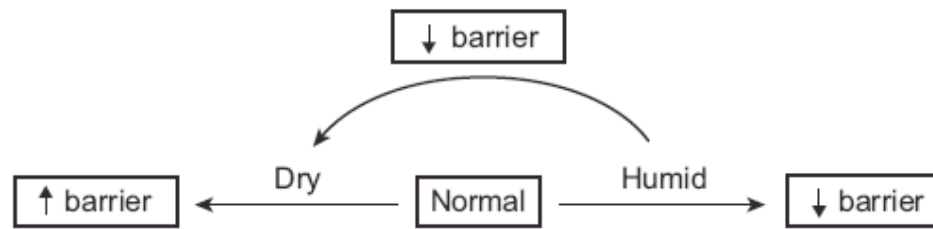


Figure 4
Changes in external humidity alter permeability barrier function.

Signals of Barrier Homeostasis

Transcriptional regulation of corneocyte protein expression by nuclear hormone receptor (NHR) ligands. Members of both the Class I and Class II family of NHR influence epidermal barrier formation, function, and development (Table II). At least three ligands (glucocorticoids, estrogens, and androgens) of the Class I family (receptors for the steroid hormones) regulate permeability barrier development in fetal skin, as well as barrier homeostasis in adult skin. In fetal skin both exogenous glucocorticoids and estrogens, either administered *in utero* or added directly to fetal skin explants in organ culture, accelerate the development of a mature permeability barrier, whereas in contrast, administration of androgens retards barrier ontogenesis (Hanley *et al*, 1996a, b, 1998). Although the impact of Class I ligands on barrier function in post-natal skin is less well understood, normal to supra-normal levels of androgens provoke an analogous decline in permeability barrier homeostasis in adult murine and human skin (Kao *et al*, 2001). Yet, an increase in endogenous glucocorticoids, induced by either psychological stress (Denda *et al*, 2000), or systemic administration of exogenous steroids (Kao *et al*, 2003), alters permeability barrier homeostasis and SC integrity/cohesion.

Table II. Cutaneous effects of class II NHR ligands

| Receptor | Ligands/activators | Fetal barrier development | Adult barrier homeostasis | Anti-inflammatory |
|---------------|--|---------------------------|---------------------------|-------------------|
| Classic | | | | |
| RAR | All-trans-retinoic acid | None | Worse | Improves |
| T3R | Triiodothyronine (T3) | Accelerates | ? | ? |
| D3R | 1,25(OH) ₂ vitamin D3 | None | Worse | Improves |
| Liposensor | | | | |
| PPAR α | Leukotriene B ₄ , fatty acids, fibrates | Accelerate | Accelerate | Improve |
| PPAR γ | Prostaglandin J ₂ , troglitazone | None | None | Improve |
| PPAR δ | Free fatty acids | None | Accelerate | Improve |
| LXR | Oxygenated sterols | Accelerate | Accelerate | Improve |

NHR, nuclear hormone receptor; PPAR, peroxisome proliferator activator receptor; LXR, liver X receptor.

Table III. Protease–anti-protease reactions in stratum corneum

| | Preferred substrate | Anti-proteases |
|---------------------|---------------------|--|
| Serine proteases | | |
| SCTE | DSG1, pro-SCCE | LEKTI 1 |
| SCCE | DSC1, CDSN | SKALP, SLPI, LEKTI 1 |
| Cysteine proteases | | |
| SCCP | DSC1, CDSN | Cystatin M/E, Cystatin α , SLPI |
| Aspartate proteases | | |
| Cathepsin G | DSC1, CDSN | SLPI |

SC, stratum corneum; SCTE, SC tryptic; SCCE, SC chymotryptic; SCCP, SC cysteine protease; DSG1, desmoglein 1; DSC1, desmocollin 1; CDSN, corneodesmosin; SLPI, secretory leukocyte protease inhibitor; SKALP, skin-derived antileukocyte proteinase.

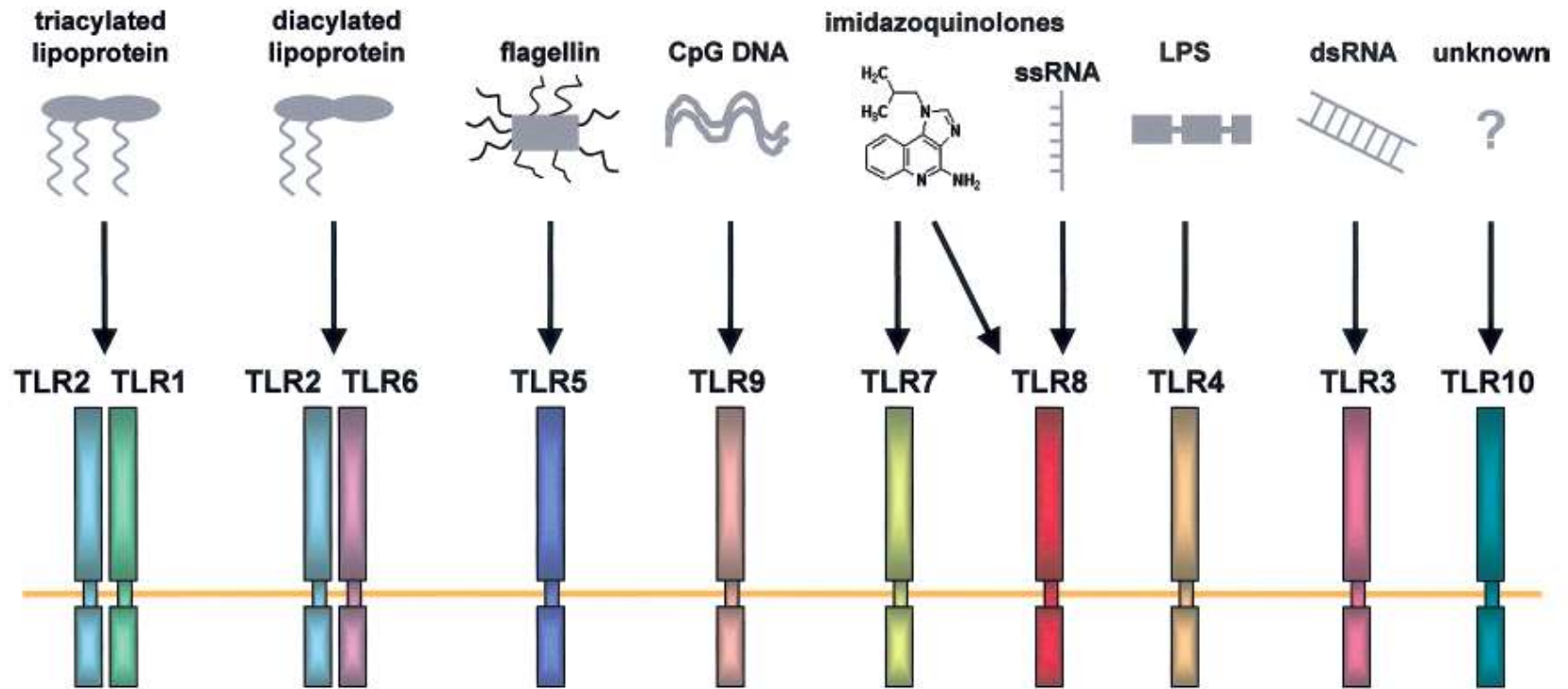


Figure 1
Human Toll-like receptors display specificity in their recognition of pathogen-associated molecular patterns and/or synthetic compounds.

Table I. Mammalian peptides and proteins relevant to skin with antimicrobial activity (AMP)^a

| | References |
|---|---|
| AMP identified in resident cells | |
| Cathelicidins | Frohm <i>et al</i> (1997), Marchini <i>et al</i> (2002) |
| β -Defensins | Harder <i>et al</i> (2001), Liu <i>et al</i> (2003) |
| Bactericidal/permeability-increasing protein (BPI) | Takahashi <i>et al</i> (2004) |
| Lactoferrin | Cumberbatch <i>et al</i> (2000) |
| Lysozyme | Marchini <i>et al</i> (2002) |
| Dermcidin | Schittek <i>et al</i> (2001), Murakami <i>et al</i> (2004) |
| RNase 7 | Harder and Schroder (2002) |
| AMP identified in infiltrating cells | |
| Cathelicidins | Gallo <i>et al</i> (1994), Marchini <i>et al</i> (2002) |
| α -Defensins | Harwig <i>et al</i> (1994) |
| Lactoferrin | Caccavo <i>et al</i> (2002) |
| Granulysin | Stenger <i>et al</i> (1998) |
| Perforin | Stenger <i>et al</i> (1998) |
| Eosinophil cationic protein (ECP)/ RNase 3 | Domachowske <i>et al</i> (1998a) |
| Eosinophil-derived neurotoxin (EDN)/RNase 2 | Domachowske <i>et al</i> (1998b) |
| Regulated upon activation, normal T cells expressed and secreted (RANTES) | Tang <i>et al</i> (2002) |

Table I. Continued

| | References |
|---|--|
| Platelet factor 4 (PF-4) | Tang <i>et al</i> (2002) |
| Connective tissue activating peptide 3 (CTAP-3) | Tang <i>et al</i> (2002) |
| Platelet basic protein | Tang <i>et al</i> (2002) |
| Thymosin β -4 (T β -4) | Tang <i>et al</i> (2002) |
| Fibrinopeptide B (FP-B) | Tang <i>et al</i> (2002) |
| Fibrinopeptide A (FP-A) | Tang <i>et al</i> (2002) |
| AMP identified as proteinase inhibitors | |
| hCAP18/LL-37 prosequence (cathelin-like domain) | Zaiou <i>et al</i> (2003) |
| Secretory leukocyte proteinase inhibitor (SLPI)/Antileukoprotease | Wingens <i>et al</i> (1998) |
| Elafin/skin-derived antileukoprotease (SKALP) | Simpson <i>et al</i> (1999), Meyer-Hoffert <i>et al</i> (2003) |
| P-cystatin α | Takahashi <i>et al</i> (1994) |
| Cystatin C | Blankenvoorde <i>et al</i> (1998) |
| AMP identified as chemokines | |
| Psoriasin | Glaser <i>et al</i> (2001) |
| Monokine induced by IFN- γ (MIG/CXCL9) | Cole <i>et al</i> (2001a) |
| IFN- γ -inducible protein of 10 kDa (IP-10/CXCL10) | Cole <i>et al</i> (2001a) |
| IFN- γ -inducible T cell α chemoattractant (I-TAC/CXCL11) | Cole <i>et al</i> (2001a) |

Antimicrobial peptides identified as neuropeptides

| | |
|---|-------------------------------|
| α -Melanocyte stimulating hormone (α -MSH) | Cutuli <i>et al</i> (2000) |
| Substance P | Kowalska <i>et al</i> (2002) |
| Bradykinin | Kowalska <i>et al</i> (2002) |
| Neurotensin | Kowalska <i>et al</i> (2002) |
| Vasostatin-1 and chromofungin (chromogranin A) | Tasiemski <i>et al</i> (2002) |
| Secretolytin (chromogranin B) | Tasiemski <i>et al</i> (2002) |
| Enkephalin and peptide B (proenkephalin A) | Tasiemski <i>et al</i> (2002) |
| Ubiquitin | Kieffer <i>et al</i> (2003) |
| Neuropeptide Y | Lambert <i>et al</i> (2002) |
| Polypeptide YY/skin-polypeptide YY | Lambert <i>et al</i> (2002) |
| Adrenomedullin | Allaker <i>et al</i> (1999) |

AMP identified based on other functions

| | |
|---|----------------------------|
| Hemoglobin-derived peptides | Parish <i>et al</i> (2001) |
| Calprotectin (MRP8/MRP14)/calgranulin A/B | Sohnle <i>et al</i> (2000) |
| Neutrophil gelatinase-associated lipocalin (NGAL) | Goetz <i>et al</i> (2002) |
| Epidermal H1 histones | Kashima (1991) |
| Myeloperoxidase | Rosen and Michel (1997) |

^aReferences limited due to space restrictions.

Table II. Mouse models demonstrate that the chemokine family is important in host defense

| Knockout | Clinical and immunological consequence Reference ^a | Microbe |
|------------------|---|-----------------------------------|
| <i>Receptors</i> | | |
| CCR1 | | |
| | Reduced inflammation, increased mortality Domachowski, J Immunol, 2000 | Paramyxovirus |
| | Increased susceptibility to infection Khan, J Immunol, 2001 | <i>Toxoplasma gondii</i> |
| | Increased susceptibility to infection Gao, J Exp Med, 1997 | <i>Aspergillus fumigatus</i> |
| | Smaller lesions containing fewer parasites Rodriguez-Sosa, Immunol Cell Biol, 2003 | <i>Leishmania major</i> |
| | No effect on corneal PMN or opacities Hall, J Immunol, 2001 | <i>Onchocerciasis</i> |
| CCR2 | | |
| | Defective macrophage recruitment and host defense Kurihara, J Exp Med, 1997 | <i>Listeria monocytogenes</i> |
| | Decreased macrophage and CD8+ T cell recruitment Huffhagle, Immunopharmacology, 2000 | <i>Cryptococcus neoformans</i> |
| | Prolonged pulmonary infection Up to 800-fold greater dissemination to spleen and brain Reduced macrophage recruitment Traynor, J Immunol, 2000 | <i>C. neoformans</i> |
| | Failure to control infection Block in infection-induced relocalization of splenic DC Sato, J Exp Med, 2000 | <i>Leishmania major</i> |
| | Significantly decreased survival Macrophages exhibit recruitment defects to lungs 100-fold higher bacterial load in lungs Peters, PNAS, 2001 | <i>Mycobacterium tuberculosis</i> |
| CCR4 | | |
| | Decreased mortality in endotoxemic shock Chvatchko, J Exp Med, 2000 | LPS |
| CCR5 | | |
| | Impaired macrophage function ANCE Reduced efficiency in bacterial clear Zhou, J Immunol, 1998 | <i>Listeria monocytogenes</i> |
| | No protection against infection or death Elvin Nature, 2004 | <i>Yersinia</i> |
| | Decreased survival Defect in leukocyte migration to brain Huffhagle, J Immunol, 1999 | <i>C. neoformans</i> |
| | Lower parasite burden in liver Sato, J Immunol, 1999 | <i>Leishmania donovani</i> |

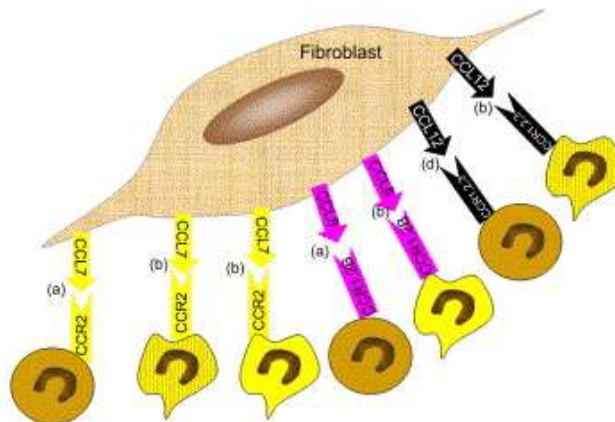
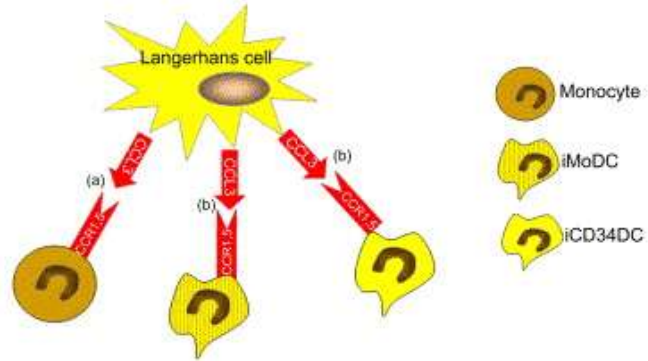
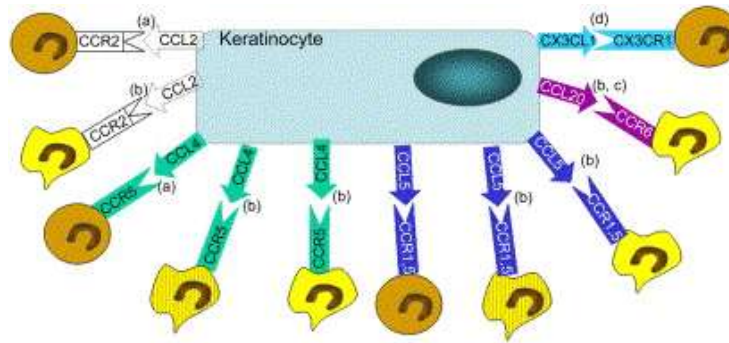
Table II. Continued

| Knockout | Clinical and immunological consequence Reference* | Microbe |
|--------------------------------------|--|--|
| | Reduced macrophage infiltration Glass, <i>Virology</i> , 2001 | Mouse hepatitis virus |
| | Antiviral T-cell response appears to be augmented Nansen, <i>Immunobiology</i> , 2002 | Lymphocytic choriomeningitis virus |
| | Decreased susceptibility to Cryptosporidiosis Campbell, <i>J Parasitol</i> , 2002 | <i>Cryptosporidium parvum</i> |
| | Decreased susceptibility to cerebral malaria Belnoue, <i>Blood</i> , 2003 | <i>Plasmodium berghei</i> |
| IL8Rα/CXCR2 | | |
| | Dysfunctional neutrophil migration Godaly, <i>J Immunol</i> , 2000 | <i>Escherichia coli</i> |
| | Subepithelial neutrophil entrapment and renal scarring Hang, <i>J Infect Dis</i> , 2000 | <i>E. coli</i> |
| | Enhanced susceptibility to pyelonephritis Freundus, <i>J Exp Med</i> , 2000 Freundus, <i>J Infect Dis</i> , 2001 | <i>E. coli</i> |
| | Impaired neutrophil recruitment Del Rio, <i>J Immunol</i> , 2001 | <i>Toxoplasma gondii</i> |
| | Impaired neutrophil extravasation Increased bacterial burden Kielian, <i>J Immunol</i> , 2001 | <i>S. aureus</i> |
| | Reduction in neutrophil recruitment Goncalves, <i>Scand J Immunol</i> , 2002 | <i>Mycobacterium avium</i> |
| | Enhanced susceptibility to herpetic stromal keratitis Banerjee, <i>J Immunol</i> , 2004 | HSV-1 |
| | Decrease in Lyme arthritis severity Brown, <i>J Immunol</i> , 2003 | <i>B. burgdorferi</i> |
| | Decreased mucus production and airway hyperreactivity Miles, <i>J Immunol</i> , 2003 | Respiratory syncytial virus |
| CXCR5 | | |
| | Accelerated transfer of intraperitoneally administered prions into the spinal cord Prinz, <i>Nature</i> , 2003 | Prions |
| DARC | | |
| | Increased inflammatory infiltrates in lung and liver Dawson TC, <i>Blood</i> , 2000 | LPS |
| Ligands | | |
| CCL2 | | |
| | Reduced NKT cell recruitment Kawakami, <i>J Immunol</i> , 2001 | <i>Cryptococcus neoformans</i> |
| | Enhanced susceptibility to gingivitis Chae, <i>Infect Immun</i> , 2002 | <i>Streptococcus mutans</i> , <i>Streptococcus intermedius</i> , <i>Peptostreptococcus micros</i> , <i>Porphyromonas gingivalis</i> , <i>Prevotella intermedia</i> , <i>Fusobacterium nucleatum</i> |

Table II. Continued

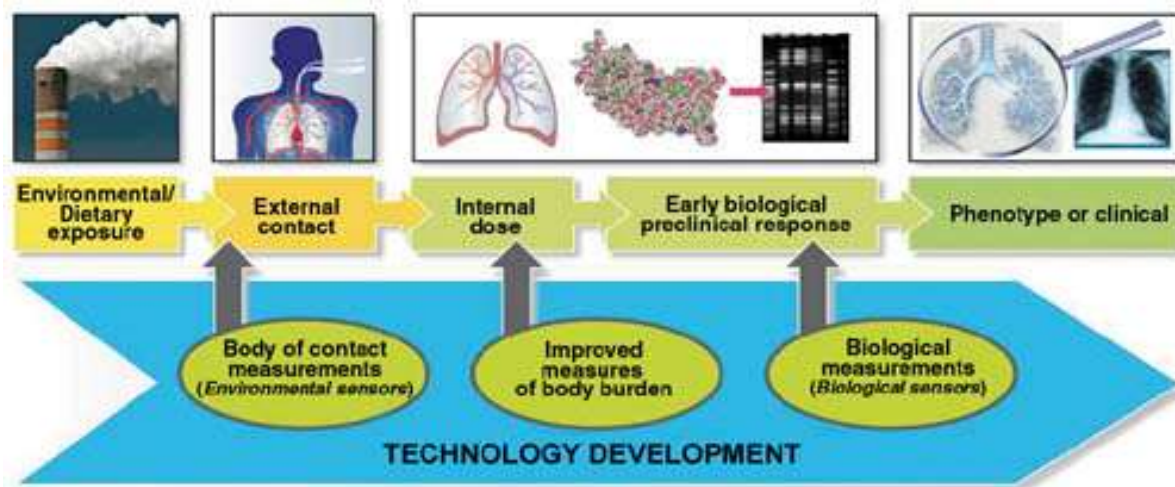
| Knockout | Clinical and immunological consequence Reference ^a | Microbe |
|---------------|--|------------------------------|
| | Failure to expel infection deSchodtmaster, J Immunol, 2003 | <i>Trichuris muris</i> |
| CCL3 | | |
| | Reduced antiviral host defense Domachowska, J Immunol, 2000 | Pneumonia virus |
| | Inhibited inflammatory and protective liver responses Salazar-Mather, J Exp Med, 1998 | Murine cytomegalovirus |
| | Decreased resistance to infection Reduced NK cell accumulation Salazar-Mather, J Clin Invest, 2000 | Murine cytomegalovirus |
| | Impaired survival Lindell, Infect Immun, 2001 | <i>Klebsiella pneumoniae</i> |
| | Decreased survival Olszewski, J Immunol, 2000 | <i>C. neoformans</i> |
| | Impaired prevention of eosinophilic pneumonia Olszewski, Infect Immun, 2001 | <i>C. neoformans</i> |
| | Reduced protective innate immunity against sepsis Cecal ligation and puncture Takahashi, J Leuk Biol, 2002 | <i>C. neoformans</i> |
| | Delayed viral clearance Trifilo, J Virol, 2003 | Mouse hepatitis virus |
| | Lower parasite burden in liver Sato, J Immunol, 1999 | <i>Leishmani donovani</i> |
| CCL11 | | |
| | Suppressed endotoxemia-associated peritoneal neutrophils Cheng, Exp Mol Pathol, 2002 | LPS |
| CXCL15 | | |
| | Impaired pulmonary host defense Chen, J Immunol, 2001 | <i>Klebsiella pneumoniae</i> |

Knockout mice (in bold) experienced improved survival advantage compared with wild-type mice.
^aAll references in this table can be found in the supplemental material available online for this article.



Balneoterapie – imunomodulační nástroj

✓ změna prostředí



Balneoterapie – imunomodulační nástroj

✓ stres

Mechanisms by Which Psychologic Stress Alters Cutaneous Permeability Barrier Homeostasis and Stratum Corneum Integrity

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Although many skin disorders, including psoriasis and atopic dermatitis, are adversely affected by psychologic stress (PS), the pathophysiologic link between PS and disease expression remains unclear. Recent studies demonstrated PS-induced alterations in permeability barrier homeostasis, mediated by increased endogenous glucocorticoids. Here, we assessed the mechanisms by which PS alters stratum corneum (SC) function. Insomniac psychologic stress (IPS) altered both barrier homeostasis and SC integrity. IPS decreased epidermal cell proliferation, impaired epidermal differentiation, and decreased the density and size of corneodesmosomes (CD), which was linked to degradation of CD proteins (e.g., desmoglein1). Barrier compromise was linked to decreased production and secretion of lamellar bodies (LB), which in turn could be attributed to a decrease in *de novo* synthesis of epidermal lipids. Topical physiologic lipids (equimolar cholesterol, ceramides, and free fatty acids) normalized both barrier homeostasis and SC integrity in IPS mice, further evidence that lipid deficiency accounted for these functional abnormalities. Thus, PS inhibition of epidermal lipid synthesis results in decreased LB formation and secretion, as well as decreased CD, compromising both permeability barrier homeostasis and SC integrity. These studies suggest that topical treatment with epidermal physiologic lipids could be beneficial in stress-induced, barrier-associated dermatoses, such as psoriasis and atopic dermatitis.

Key words: corneodesmosome/epidermal lipid synthesis/lamellar body/stratum corneum integrity/trans-epidermal water loss

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